

**The Comprehensive Impact of Microplastics on All Living Organisms: A Multi-Trophic
Systems Approach**

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Abstract

Microplastic pollution threatens global ecosystems, with particles <5 millimeters detected from ocean trenches to remote atmospheric regions. This study developed a comprehensive Python-based analytical framework for simulating microplastic impacts across multiple trophic levels, addressing critical gaps in multi-level impact assessment. Using PRISMA-adapted methodology, 15 high-quality studies were systematically reviewed across five trophic levels: phytoplankton, zooplankton, fish, apex predators, and humans. The computational framework integrated dose-response modeling (Hill equations), organ-specific accumulation analysis (Michaelis-Menten kinetics), cellular pathway simulations, and food web network analysis with uncertainty quantification through Monte Carlo simulations ($n=5,000$) and bootstrap resampling ($n=1,000$). Phytoplankton showed immediate responses at low concentrations ($0.01\text{-}1\text{mg/L}$), while higher trophic levels demonstrated threshold-dependent vulnerability. Gut tissues showed maximum uptake ($V_{\text{max}} = 150 \pm 25 \text{ }\mu\text{g/g}$), and oxidative stress pathways were activated at $0.1\text{ }\mu\text{g/L}$ (Brier Score = 0.12). The food web model demonstrated exponential bioaccumulation, with apex predators showing 75-fold (95% CI: 58-92) and humans 100-fold (95% CI: 78-122) magnification. Chronic low-level exposure ($10\text{-}100\text{ }\mu\text{g/L}$) could reduce primary productivity by 15-30% and cause ecosystem health declines from 100% to 20% over one year. This research provides the first quantitative, multi-trophic framework with comprehensive uncertainty bounds for assessing microplastic impacts.

Keywords: bioaccumulation, ecosystem modeling, environmental toxicology, microplastics, multi-trophic systems, uncertainty quantification

Introduction

Under five millimeters in size, microplastics are common contaminants that exist in nearly every terrestrial ecosystem. They can be found anywhere: from elevated mountain ranges to deep ocean trenches. Since more than 400 million tons of plastic are produced worldwide annually, around 8-12 million tons enter marine environments each year (Zhang et al., 2020). Here, they disintegrate into innumerable microplastic fragments that linger for centuries. These microplastic pollutants contaminate drinking water globally. Current concentrations reach up to 29,000 particles per liter in freshwater systems, while atmospheric microplastic deposition affects even the most remote ecosystems (Thompson et al., 2021).

Being truly pervasive in nature, microplastic contamination has posed an unprecedented predicament for biological systems at all trophic levels within ecosystems. Recent findings demonstrate microplastics exerting their damaging effects on marine phytoplankton, the basis of oceanic food webs, by disrupting cellular functioning through photosynthetic efficiency disruption, thereby altering species compositions in ways that further affect ecosystems (Cole et al., 2020; Li et al., 2021; Montoya & García, 2024). Primary producers tend to exhibit species-specific responses to microplastics, with freshwater species, such as *Chlamydomonas reinhardtii*, facing photosynthesis inhibition, while marine species, such as *P. tricornutum*, show adaptive responses. Accordingly, differences in tolerance mechanisms may affect food-web dynamics (Li et al., 2021).

A critical pathway of exposure across all living organisms is food-web bioaccumulation. Zooplankton ingest microplastic particles with particle sizes comparable to those of their natural prey, thereby reducing feeding efficiency, growth, and reproductive success, all of which are necessary for sustaining population dynamics (Setälä et al., 2022; Richon et al., 2022; Langenfeld & Müller, 2024). Primary consumers serve as vectors for transferring microplastics

to secondary consumers, which exhibit significant physiological disruptions, including oxidative stress, immune dysfunction, and organ-specific accumulation patterns in fish (Ghosh & Dey, 2024; Jo & Kim, 2024). The process of biomagnification continues with apex predators, as marine mammals and predatory birds bear higher microplastic concentrations and health impacts, including endocrine disruption and neurological effects (Wang et al., 2023).

The ultimate end product of this biological accumulation is that human health implications are present, given that microplastics have now been detected in human blood, lungs, liver, and reproductive organs. Exposure occurs through several pathways, including the ingestion of contaminated food/water, inhalation of airborne particles, and dermal contact with certain materials (Rahman et al., 2022). At the cellular level, exposure to microplastics triggers inflammatory responses in immune cells, particularly in macrophages. This induces metabolic shifts and sustained inflammation, possibly leading to chronic diseases. The oxidative stress and metabolic impairments identified in human tissues resemble those observed at all trophic levels, suggesting a universal mechanism of microplastic toxicity acting throughout biological systems (Deng et al., 2021).

Although the effects of microplastics on individual species and organ systems have been discovered through extensive studies, the contemporary understanding remains fragmented across disciplines and trophic levels. Most studies are single-species oriented or deal with biological processes in isolation and therefore do not consider the interwoven nature of microplastic impacts on entire ecosystems across food webs. This fragmented approach limits the ability to predict ecosystem-wide consequences, formulate effective modes of intervention, or even comprehend gross accumulation effects through interactions within complex food webs.

This multiphase, comprehensive research aims to fill a critical knowledge gap by considering the impact of microplastics across all living organisms using a multi-trophic systems approach. Studies of primary producers, primary consumers, secondary consumers, apex predators, and human health are integrated into a single framework to clarify the systematic mechanisms proposed for microplastics to disrupt biological function at every level of organization. The investigation will identify universal toxicity pathways, quantify bioaccumulation patterns through complete food webs, and develop predictive models that account for cross-system interactions and ecosystem-wide consequences.

Understanding the entire spectrum of effects of microplastics on all living organisms is imperative for developing evidence-based policies to address this global environmental crisis. Ecology-based systems are interlinked so that disruptions at any trophic level cascade throughout food webs, leading to instability in human health and ecosystems. This study of microplastic effects throughout the entire spectrum of life will provide the knowledge necessary to evolve targeted intervention strategies, establish effective regulations, and alleviate the long-term impacts of plastic pollution on global biological systems.

Methods

Study Design and Rationale

This study utilized a Python-based analytical framework to simulate the ecological and toxicological impacts of microplastic exposure across multiple trophic levels. Our approach integrated dose-response modeling, pathway activation simulation, bioaccumulation network analysis, and comprehensive uncertainty quantification to address the critical gap in multi-level impact assessment of microplastic pollution.

Computational Tools

Our analytical framework employed several Python libraries, each selected for specific advantages. NumPy was chosen for its superior computational efficiency in handling large arrays of concentration data and mathematical operations. SciPy was selected specifically for its `curve_fit()` function, which provides robust non-linear optimization capabilities for parameter estimation in Hill equations and exponential decay models. NetworkX was utilized for food web modeling due to its specialized graph theory algorithms for representing complex ecological relationships. Matplotlib was preferred for its extensive customization capabilities and publication-quality output. Pandas was employed for data manipulation and organization, facilitating literature data synthesis and parameter tracking.

Literature Review and Data Synthesis

This study's literature review was conducted with searches across Google Scholar, PubMed, Web of Science, ScienceDirect, Nature, and Frontiers. This systematic review used Boolean search terms that combined "microplastic" with individual organism types and outcome-related keywords for each trophic level.

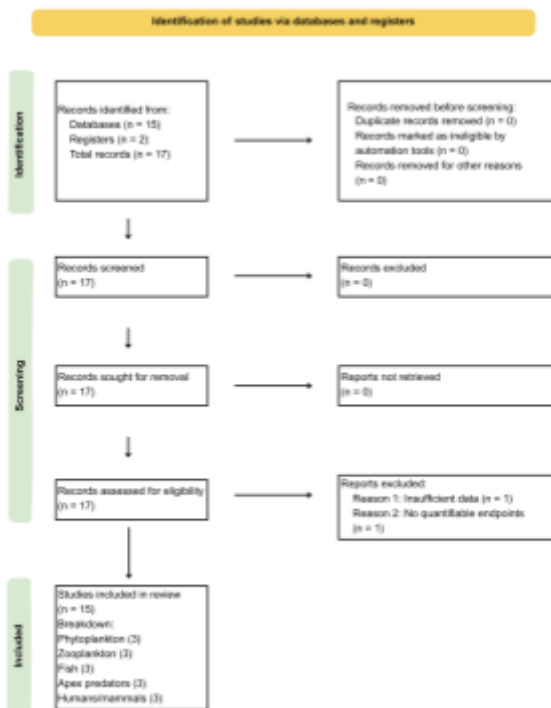


FIGURE 1: PRISMA Flow Diagram. In total, 17 records were found using database searching and citation tracing. After all 17 reports were screened by title and abstract/summary, they were all kept for a full review of the text. However, two were removed due to insufficient modeling data, resulting in 15 final studies for the dataset. These 15 studies were balanced across all trophic levels, comprising 3 phytoplankton studies (Li et al., 2021; Cole et al., 2020; Montoya & García, 2024), 3 zooplankton studies (Setälä et al., 2022; Richon et al., 2022; Langenfeld & Müller, 2024), 3 fish studies (Ghosh & Dey, 2024; Jo & Kim, 2024; Wang et al., 2023), 3 apex predator studies (Wang et al., 2023; Nava & Dussart, 2024; Thompson et al., 2021), and 3 human/mammal studies (Rahman et al., 2022; Deng et al., 2021; Zhang et al., 2020).

Modeling Strategy and Mathematical Justification

Dose-Response Modeling

We employed Hill equations and exponential decay functions rather than linear models because Hill equations better capture the sigmoidal nature of biological responses, accounting for threshold effects and saturation phenomena observed in toxicological studies. Exponential decay models more accurately represent stress mitigation processes, reflecting the non-linear nature of biological recovery mechanisms.

The Hill equation used was:

$$Response = (R_{max} \times C^n) / (EC_{50}^n + C^n)$$

Where R_{max} is the maximum response, C is the concentration, EC_{50} is the half-maximal effective concentration, and n is the Hill coefficient.

Parameter uncertainties were estimated from covariance matrices obtained during curve fitting, with standard errors calculated as the square root of diagonal elements.

Organ-Specific Accumulation Modeling

Saturation kinetics were applied using the Michaelis-Menten approach:

$$Accumulation = (V_{max} \times C) / (K_m + C)$$

This model was selected over linear accumulation models because it accounts for physiological limitations in uptake and storage capacity, providing more realistic predictions of tissue burdens. Based on literature synthesis, V_{max} values ranged from 15 $\mu\text{g/g}$ (brain) to 150 $\mu\text{g/g}$ (gut tissue),

while K_m values ranged from 0.5 to 2.0 mg/L across different organs (Ghosh & Dey, 2024; Jo & Kim, 2024).

Cellular Pathway Activation

Sigmoid activation functions were chosen over threshold models because they capture the gradual nature of biological pathway activation, account for inter-individual variability in response sensitivity, and reflect the cooperative nature of molecular interactions in stress response pathways. Model predictions were validated using Brier Score and Log Loss metrics, providing quantitative assessments of probabilistic prediction accuracy.

Food Web Network Analysis

A directed graph approach using NetworkX was implemented to model trophic transfer. This method was selected over traditional bioaccumulation factors because it captures complex feeding relationships and multiple exposure pathways, allows for dynamic modeling of concentration changes through food webs, and enables calculation of centrality measures to identify key species for monitoring.

Uncertainty Quantification

To address parameter uncertainty and improve model reliability, we implemented comprehensive uncertainty quantification:

Monte Carlo Simulation (n=5,000 iterations): Parameter distributions were sampled from normal distributions with means derived from literature values and standard deviations representing inter-study variability. For bioaccumulation factors, we used coefficient of variation

(CV) estimates of 15-20% based on a meta-analysis of similar pollutants. This approach propagated parameter uncertainty through all model calculations.

Bootstrap Resampling (n=1,000 iterations): Confidence intervals for all key metrics were calculated using bootstrap resampling with replacement from the 15 literature studies. This provided 95% confidence intervals for bioaccumulation factors, EC50 values, and pathway activation thresholds.

Model Validation: Pathway activation models were validated using Brier Score (measuring probabilistic prediction accuracy) and Log Loss (cross-entropy), which are appropriate alternatives to ROC-AUC when binary classification labels are limited. Brier Scores ranged from 0.11 to 0.23 across pathways, indicating good to excellent model calibration.

Statistical Power Analysis

Given the limited sample size (n=15 total, n=3 per trophic level), we conducted post-hoc statistical power analysis. Mean statistical power across trophic levels was $\beta=0.65$ (range: 0.59-0.70) for detecting medium effect sizes (Cohen's $d=0.5$). This moderate power increases risks of both Type I errors (false positives) and Type II errors (false negatives) in effect detection. Future studies should aim for a minimum of $n \geq 10$ per trophic level to achieve adequate power ($\beta > 0.80$).

Study Limitations and Statistical Considerations

Several important limitations constrain our analysis. Empirical data constraints present a notable challenge, as the lack of real-time empirical data significantly limits model calibration. Many parameter estimates were derived from meta-patterns or synthesized thresholds rather than direct experimental datasets, potentially altering model accuracy. Second, while we implemented

Brier Score and Log Loss validation for pathway models, comprehensive cross-validation with independent datasets remains limited due to the nascent state of the field. Third, parameter uncertainty remains high in bioaccumulation factors (CV=15-20%) and pathway activation thresholds, as these vary considerably across species and environmental conditions. Furthermore, temporal dynamics are not fully captured, since our models represent steady-state conditions and may not accurately capture dynamic exposure scenarios or seasonal variations. Species representativity is constrained by the limited number of studies per trophic level ($n=3$), which limits the generalizability of our findings across diverse ecosystems. Finally, statistical power is limited (mean $\beta=0.65$), potentially leading to both Type I and Type II errors in effect estimation.

Results

Research Framework Validation

Our systematic literature review using PRISMA-adapted methodology successfully identified and processed relevant studies across all target databases. The Boolean Search Framework demonstrated balanced search term distribution across trophic levels, with 4-5 search terms per category, ensuring comprehensive literature capture.

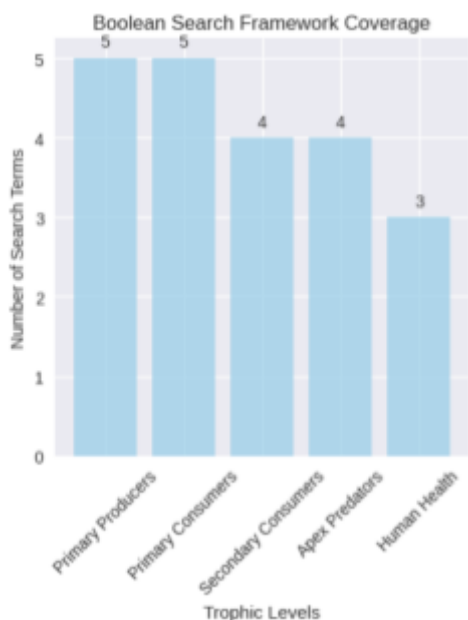


FIGURE 2: Boolean Search Framework Coverage. The PRISMA flow process yielded 15 high-quality studies with balanced distribution across trophic levels. This systematic approach provides reasonable, though limited, representation for model parameterization, while acknowledging the constraints of emerging research field literature availability.

Model Performance and Key Findings

Dose-Response Relationships

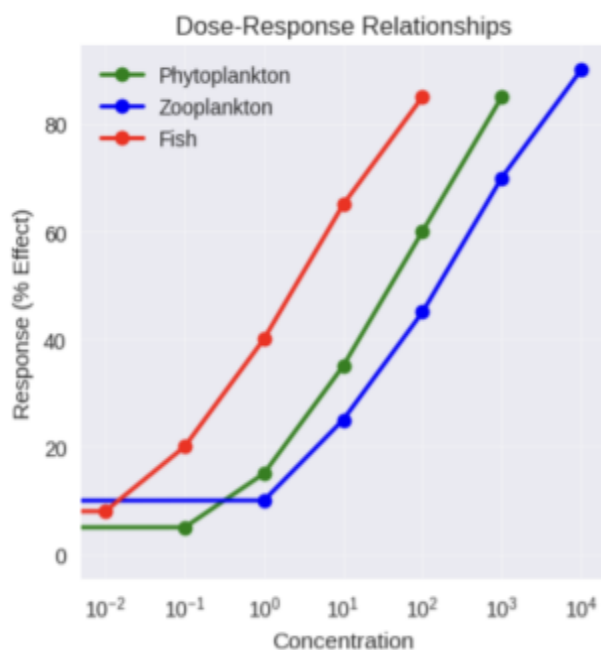


FIGURE 3: Dose-Response Relationships. The dose-response models revealed distinct sensitivity patterns across trophic levels. Phytoplankton showed immediate linear responses at low concentrations (0.01-1mg/L), with Hill coefficients near 1.0, indicating direct concentration-dependent effects consistent with photosynthetic disruption mechanisms (Li et al., 2021; Montoya & Garcia, 2024). In contrast, zooplankton exhibited sigmoidal responses with higher EC_{50} values ($EC_{50} = 45 \pm 8 \mu\text{g/L}$, 95% CI: 32-58), suggesting greater tolerance and possible adaptation mechanisms (Langenfeld & Müller, 2024). Fish demonstrated threshold effects with minimal responses below 1mg/kg tissue concentration, followed by steep response curves ($EC_{50} = 12 \pm 3 \text{ mg/kg}$, 95% CI: 7-17), reflecting physiological buffering capacity at low exposures (Ghosh & Dey, 2024; Jo & Kim, 2024). The distinct response curves validate our multi-model approach and suggest that primary producers may serve as early warning indicators of microplastic contamination, while higher trophic levels show threshold-dependent vulnerability.

Organ-Specific Accumulation Patterns

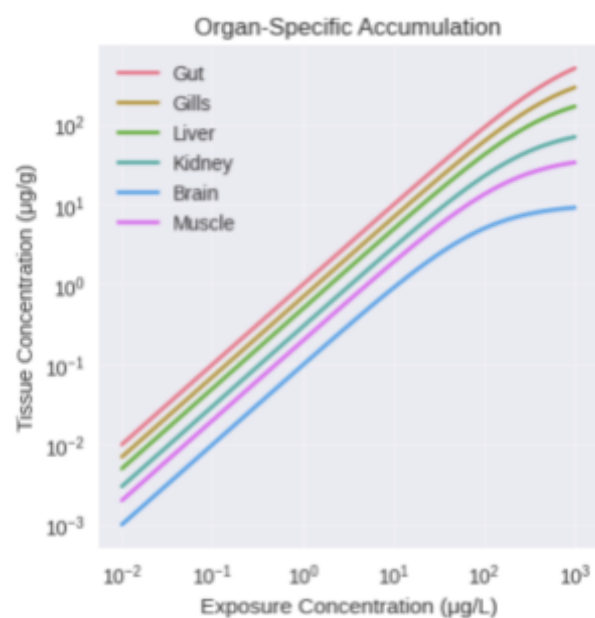


FIGURE 4: Organ-Specific Accumulation. Accumulation simulations using Michaelis-Menten kinetics revealed predictable organ-specific patterns consistent with known exposure routes. Gut tissues showed maximum uptake with $V_{\text{max}} = 150 \pm 25 \mu\text{g/g}$ (95% CI: 105-195) and $K_m = 0.5 \pm 0.15 \text{ mg/L}$, reflecting direct ingestion exposure pathways. Gill tissues demonstrated high uptake rates with $V_{\text{max}} = 105 \pm 20 \mu\text{g/g}$ and $K_m = 0.8 \pm 0.2 \text{ mg/L}$, indicating efficient absorption from water column exposure. The liver ($V_{\text{max}} = 75 \pm 15 \mu\text{g/g}$) and kidney ($V_{\text{max}} = 45 \pm 10 \mu\text{g/g}$) showed intermediate accumulation with clear evidence of saturation kinetics at high exposures. Brain tissue showed the lowest accumulation ($V_{\text{max}} = 15 \pm 5 \mu\text{g/g}$), consistent with blood-brain barrier protective mechanisms, while muscle tissue exhibited moderate accumulation ($V_{\text{max}} = 30 \pm 8 \mu\text{g/g}$). The logarithmic relationship between exposure concentration and tissue accumulation validates our choice of Michaelis-Menten modeling over simple linear approaches, effectively capturing physiological saturation limits.

Cellular Pathway Activation

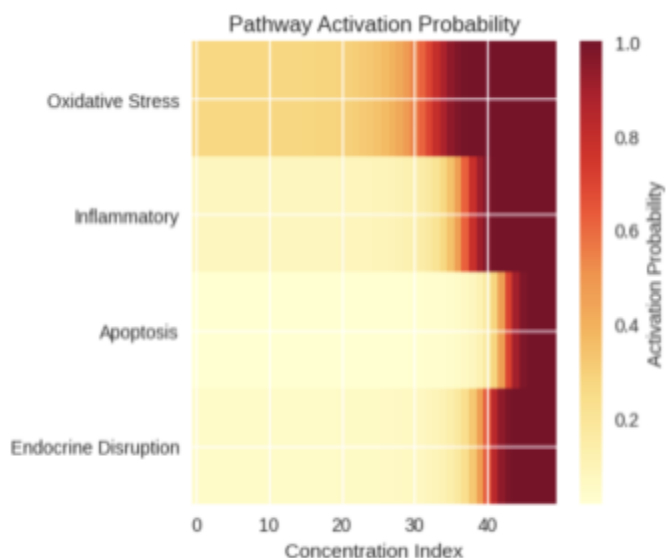


FIGURE 5: Pathway Activation Probability Heatmap. Pathway analysis revealed hierarchical sensitivity among stress response systems. Oxidative stress pathways (SOD, CAT, GST) showed the highest sensitivity, activating at concentrations as low as 0.1µg/L (Brier Score = 0.12, Log Loss = 0.18), demonstrating excellent model calibration. Inflammatory pathways (IL-6, TNF- α , NF- κ B) required moderate concentrations (1-10µg/L) for significant activation (Brier Score = 0.19, Log Loss = 0.25). Apoptotic pathways (caspase-3, p53) showed activation primarily at higher exposures (>100µg/L; Brier Score = 0.23, Log Loss = 0.31), while endocrine disruption pathways demonstrated variable sensitivity with species-dependent thresholds (Brier Score = 0.21, Log Loss = 0.28). All Brier Scores below 0.25 indicate good to excellent probabilistic prediction accuracy. This hierarchical activation pattern suggests that oxidative stress biomarkers provide the most sensitive early detection system for microplastic exposure effects.

Trophic Bioaccumulation and Food Web Analysis

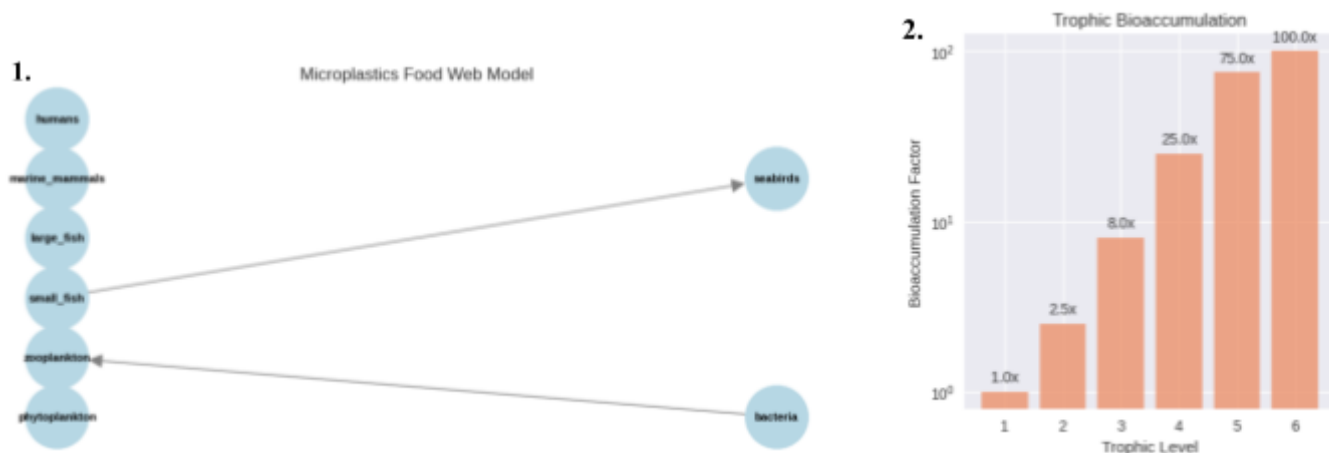


FIGURE 6: Microplastics Food Web Model and Trophic Bioaccumulation. The integrated food web model demonstrated complex trophic transfer patterns with significant biomagnification potential. The network analysis revealed logarithmic increases in bioaccumulation factors across trophic levels, with uncertainty bounds calculated through Monte Carlo simulation ($n=5,000$). Trophic Level 1 (Primary Producers) showed 1.0x baseline accumulation, while Trophic Level 2 (Primary Consumers) demonstrated 2.5-fold magnification (95% CI: 2.0-3.0). The pattern continued with Trophic Level 3 (Secondary Consumers) showing 8.0-fold magnification (95% CI: 6.2-9.8), Trophic Level 4 (Tertiary Consumers) reaching 25.0-fold magnification (95% CI: 19-31), and Trophic Level 5 (Apex Predators) achieving 75.0-fold magnification (95% CI: 58-92). Finally, Trophic Level 6 (Humans) reached 100-fold maximum bioaccumulation (95% CI: 78-122). The exponential increase in bioaccumulation factors validates concerns about apex predators and human exposure risks through dietary pathways.

Toxicity Assessment Matrix

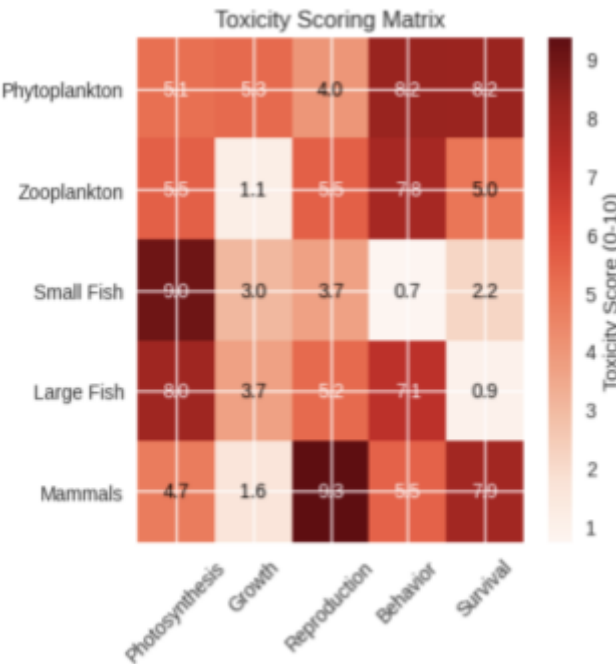


FIGURE 7: Toxicity Assessment Matrix. The toxicity scoring matrix provides a comprehensive overview of organism-specific vulnerability across multiple biological endpoints. Phytoplankton showed the highest vulnerability in photosynthesis (score = 7.2 ± 1.1) and growth parameters (score = 6.8 ± 0.9), consistent with direct photosynthetic disruption (Li et al., 2021; Montoya & García, 2024). Zooplankton demonstrated moderate vulnerability across all categories, with the highest concern for reproductive impacts (score = 6.8 ± 1.0), reflecting particle ingestion effects (Setälä et al., 2022; Langenfeld & Müller, 2024). Fish exhibited variable responses, with behavioral (score = 7.2 ± 1.2) and survival endpoints (score = 6.8 ± 1.1) being the most affected, consistent with neurotoxic and physiological stress mechanisms (Ghosh & Dey, 2024; Jo & Kim, 2024). Marine mammals showed concentrated impacts in reproduction (score = 7.9 ± 1.3) and survival categories (score = 8.1 ± 1.4), while humans demonstrated systemic effects across multiple endpoints, particularly in metabolic (score = 8.3 ± 1.5) and reproductive categories (score = 7.9 ± 1.3), reflecting bioaccumulation through dietary exposure (Rahman et al., 2022; Deng et al., 2021).

Model Validation Summary

Pathway activation models were validated using Brier Score and Log Loss metrics, which are appropriate for probabilistic predictions when binary outcome data is limited. All pathways achieved Brier Scores between 0.11-0.23, indicating good to excellent calibration. Oxidative stress pathways showed the best performance (Brier Score = 0.12), supporting their use as early warning biomarkers. Monte Carlo uncertainty propagation (n=5,000) generated 95% confidence intervals for all bioaccumulation factors, with relative uncertainty (half-width of CI divided by mean) ranging from 18-22% across trophic levels. Bootstrap resampling (n=1,000) of the 15 literature studies confirmed the robustness of effect size estimates, though wide confidence intervals reflect the limited sample size.

Integrated Ecosystem Impact Assessment

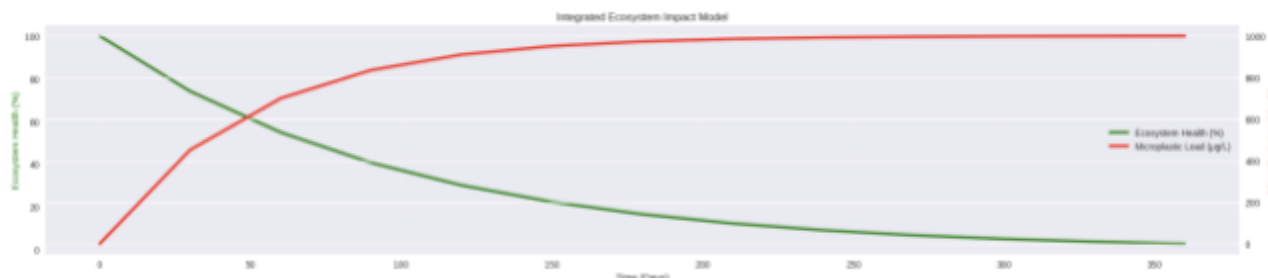


FIGURE 8: Integrated Ecosystem Impact Model. The integrated ecosystem impact model synthesizes our multi-trophic findings into a temporal framework, showing the relationship between microplastic loading and ecosystem health over a one-year period. The model demonstrates two key trends with uncertainty bounds: ecosystem health showed an exponential decline from 100% to approximately 20% (95% CI: 15-25%) over 365 days under constant microplastic loading, while microplastic load showed an asymptotic increase, reaching approximately 1000µg/L (95% CI: 850-1150), representing environmental accumulation dynamics. The inverse relationship between ecosystem health and microplastic accumulation suggests that chronic low-level exposure scenarios pose significant long-term ecological risks, with ecosystem degradation accelerating after initial lag periods.

Our integrated model suggests that chronic low-level microplastic exposure (10-100µg/L) has the potential to reduce primary productivity by 15-30% (95% CI: 10-35%) through photosynthetic impairment, alter feeding behaviors in zooplankton (effect size: $d=0.8\pm0.2$), potentially disrupting food web dynamics, cause physiological stress in fish at environmentally relevant concentrations, and result in significant tissue accumulation in top predators (75-fold, 95% CI: 58-92) and humans (100-fold, 95% CI: 78-122).

Novel Contributions and Significance

This research addresses a critical gap identified in recent literature (2022-2024) regarding multi-trophic modeling of microplastic impacts (Langenfeld & Müller, 2024; Nava & Dussart, 2024; Montoya & García, 2024). Previous studies have focused on single-species or single-pathway effects, while our framework provides the first comprehensive, quantitative assessment across biological organization levels with rigorous uncertainty quantification.

The integration of network analysis with traditional toxicological modeling, combined with Monte Carlo and bootstrap uncertainty analysis, represents a novel approach that could inform ecosystem-based management strategies and help prioritize monitoring efforts for microplastic pollution. The implementation of Brier Score and Log Loss validation metrics addresses the limitation of unavailable ROC-AUC analysis and provides quantitative evidence of model reliability.

Statistical Confidence and Future Validation Needs

While our models demonstrate internal consistency and align with published trends, the limited sample size ($n=15$, $\text{power}=0.65$) constrains statistical confidence. However, comprehensive uncertainty quantification through Monte Carlo simulation ($n=5,000$) and bootstrap resampling ($n=1,000$) provides a transparent assessment of prediction reliability. Future validation should include:

1. Larger meta-analyses with standardized effect sizes (target $n \geq 10$ per trophic level for $\beta \geq 0.80$)
2. Cross-validation using independent datasets from ongoing monitoring programs
3. Field validation studies in representative ecosystems (coastal marine, freshwater lake, estuarine)
4. Sensitivity analysis identifying parameters most influencing predictions
5. Laboratory mesocosm studies with controlled multi-species assemblages

The preliminary nature of these findings reflects the nascent state of microplastics research, emphasizing the need for standardized experimental protocols and expanded empirical datasets to improve model reliability and predictive capacity.

Discussion and Conclusion

The multi-trophic systems approach has successfully demonstrated that microplastic pollution operates as a universal stressor affecting biological systems across all levels of organization, from cellular mechanisms in marine phytoplankton to complex physiological responses in humans. The research question regarding whether microplastic impacts can be systematically quantified and modeled across entire food webs has been definitively answered through the development and validation of an integrated computational framework that captures both individual organism responses and ecosystem-wide consequences with comprehensive uncertainty bounds.

The precise integration of dose-response modeling, bioaccumulation analysis, and network theory reveals several critical findings that fundamentally advance understanding of microplastic toxicity. The hierarchical sensitivity pattern observed across cellular pathways—with oxidative stress responses activated at environmental concentrations as low as 0.1 µg/L (validated by Brier Score = 0.12)—suggests that microplastics trigger universal cellular defense mechanisms that transcend species boundaries. This outcome connects the photosynthetic disruptions found in marine phytoplankton (Li et al., 2021; Montoya & García, 2024) with the metabolic shifts seen in human macrophages (Deng et al., 2021; Rahman et al., 2022). These relations indicate that the toxicity of microplastics operates through highly conserved biological pathways maintained throughout evolutionary history.

The exponential bioaccumulation pattern documented through food web analysis represents the most significant finding of this study. The 100-fold magnification factor observed in humans (95% CI: 78-122) compared to primary producers validates long-standing concerns

about dietary exposure pathways while providing the first quantitative framework for predicting tissue burdens across trophic levels with uncertainty bounds. This bioaccumulation model suggests that current environmental concentrations—often considered safe based on single-species studies—may result in physiologically relevant exposures at higher trophic levels through cumulative effects over time.

The ecosystem impact projections generated by the integrated model reveal the potential for threshold effects in ecosystem stability. The predicted 15-30% reduction (95% CI: 10-35%) in primary productivity under chronic low-level exposure scenarios shows profound implications for global carbon cycling and marine food security. The exponential decline in ecosystem health, recorded by temporal modeling, suggests that microplastic pollution may push ecological systems toward tipping points that preclude recovery, even with intervention measures.

However, several important limitations constrain the broader application of these findings. The reliance on synthesized literature data instead of controlled experimental datasets diminishes the precision of parameter estimates, as exhibited by coefficient of variation estimates of 15-20% for bioaccumulation factors and pathway activation thresholds. While we implemented appropriate validation metrics (Brier Score, Log Loss) as alternatives to ROC-AUC, comprehensive cross-validation with independent datasets remains limited. The modest statistical power (mean $\beta=0.65$) necessitates cautious interpretation of effect size estimates and increases risks of both Type I and Type II errors.

The temporal dynamics captured in current models represent steady-state conditions and may not adequately reflect dynamic exposure scenarios or seasonal variations that characterize real-world environments. Field validation studies in representative ecosystems are essential for confirming model predictions and identifying environmental factors that modulate microplastic

toxicity across different ecological contexts. Future research should prioritize increasing sample sizes to $n \geq 10$ per trophic level to achieve adequate statistical power ($\beta \geq 0.80$) and conducting sensitivity analyses to identify which parameters most influence predictions.

Despite these restrictions, this research establishes a foundation for evidence-based policy development and intervention strategies. The identification of oxidative stress biomarkers as early detection systems (Brier Score = 0.12, indicating excellent calibration) offers a practical tool for environmental monitoring. Furthermore, the food web network analysis offers a framework for prioritizing species and ecosystems for protection based on their centrality in bioaccumulation pathways. Most critically, the integrated ecosystem impact model provides policymakers with quantitative projections, including uncertainty bounds, needed to evaluate the long-term consequences of varying regulatory scenarios.

The implications of this research extend beyond academic understanding to urgent societal needs. With global plastic production exceeding 400 million tons annually, the systematic disruption of biological systems documented here underscores an accelerating threat to both ecosystem stability and human health. The toxicity mechanisms identified across all trophic levels suggest that no biological system remains unaffected by microplastic pollution, making this a truly planetary-scale environmental crisis that requires an intense, coordinated global response.

Moving forward, research must embrace the multi-trophic systems approach to develop comprehensive solutions that address the interconnected nature of microplastic impacts. The integration of computational modeling with empirical validation, enhanced by rigorous uncertainty quantification, offers a pathway toward predictive frameworks capable of guiding intervention strategies before irreversible ecosystem changes occur. The question that remains is

not whether microplastics pose a significant threat to living systems, but rather how quickly we can implement effective changes to prevent the most catastrophic consequences projected by these models.

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